Terpenoids from Salvia kronenburgii

Gülaçtı Topçu^{*,†,‡} and Ayhan Ulubelen[‡]

TUBITAK, Marmara Research Center, Department of Chemistry, P.O. Box 21, 41470 Gebze-Kocaeli, Turkey, and Faculty of Pharmacy, University of Istanbul, 34452 Istanbul, Turkey

Received April 16, 1999

A new norditerpenoid and six new triterpenoids were isolated from the roots of Salvia kronenburgii. The new diterpenoid was kronenquinone (1). The new triterpenoids were 1β , 2α , 3β , 11α -tetrahydroxy-olean-12-ene (2), 1β , 2α , 3β , 11α -tetrahydroxy-urs-12-ene (3), 2α -acetoxy-urs-5, 12-diene- 3β , 11α -diol (4), 3α -acetoxyurs-12-ene $(\mathbf{6})$, 2α , 3β , 11α -trihydroxy-urs-12-ene ($\mathbf{6}$), 2α , 3β , 11α -trihydroxy-urs-12-ene ($\mathbf{7}$), and the known triterpenes 3β -acetoxy-urs-12-ene- 1β , 2α , 11α -triol (8) and 3β -acetoxy-olean-12-ene- 1β , 2α , 11α triol (9). Structures were determined by NMR and MS techniques.

There are about 500 Salvia species grown naturally in the world. Turkey has 90 species, 44 of them endemic. Until now, we have investigated 35 Salvia species¹⁻⁵ and isolated more than 50 triterpenoids and about 150 diterpenoids as well as some flavonoids. When we studied the biological aspects of these diterpenoids we found antibacterial⁴ and antitubercular² activities. In continuation of our studies on Turkish Salvia species, we have now studied the roots of an endemic plant Salvia kronenburgii Rech. (Lamiaceae) and have isolated nine known diterpenoids: taxodione,⁶ ferruginol,⁷ salvipisone,⁸ 12-hydroxy-sapriparaquinone,⁵ royleanone, 7-acetoxyroyleanone,⁹ 6-hydroxysalvinolone, 1-oxosalvibretol,¹⁰ and Δ^7 -manool¹¹ and a new norditerpenoid, kronenquinone (1). The same extract has also yielded eight triterpenoids, six of which are new compounds (2-7).

Results and Discussion

The molecular formula of the new norditerpene kronenquinone (1), C₁₉H₁₈O₅, indicated 11 double bond equivalents, of which three were accounted for by a tricyclic skeleton, two by quinoid oxo groups, and six by double bonds. The ¹H NMR spectrum exhibited four methyl signals; two of them, at δ 2.54 and 2.65, indicated two aromatic methyl groups. Two other methyl groups, observed at δ 1.32 and 1.33 (Me-16 and Me-17), together with the signal at δ 3.38, indicated the presence of an isopropyl group. Two aromatic protons were observed at δ 7.18 and 7.64. The UV spectrum exhibited maxima at 395 and 280 nm, indicating an extended unsaturated structure. The IR spectrum indicated a paraquinoid ring system, supported by ¹³C NMR signals at δ 182.8 and 184.2. The ¹³C NMR (APT) spectrum exhibited four CH₃, three CH, and 12 quaternary carbon signals. There were five oxygen functions in the molecule, two accounted for by the paraquinoid group. The remaining three oxygens were hydroxy groups attached to carbons whose signals appeared at δ 135.7, 150.5, and 153.2.

In a previous study with *S. multicaulis*² we isolated two fully aromatic and two orthoquinoid diterpenoids with spectra similar to those of compound 1. The HETCOR experiment showed a direct relation between protons and carbons, and the unambiguous structure of 1 was deduced by a COLOC experiment giving correlations between C-1



OН

 $R_1 = R_2 = R_4 = OH, R_3 = \beta \cdot OAc$ 8

and H-18, C-2 with H-18 and H-19, C-5 with H-7 and H-19, C-6 with H-10, C-8 with H-6 and H-7, C-9 with H-7, C-10 with H-6, C-12 with H-15, C-13 with H-16 and H-17, C-14 with H-7, and C-15 with H-16 and H-17. Thus, the structure of kronenquinone (1) was established as 1,2,12trihydroxy-11,14-dioxo-abieta-1,3,5(10),6,8,12-hexaene.

The isolated triterpenoids appeared to have either olean-(2, 7, 9) or ursane-(3, 4, 5, 6, 8) type skeletons from their

10.1021/np990165p CCC: \$18.00

© 1999 American Chemical Society and American Society of Pharmacognosy

Published on Web 10/26/1999

^{*} To whom correspendence should be addressed. Tel.: 0090 (212) 514 03 55. Fax: 0090 (212) 519 30 86. E-mail: topcu@mam.gov.tr. [†] TUBITAK, Marmara Research Center.

[‡] Faculty of Pharmacy, University of Istanbul.

Table 1.	¹ H NMR	Data (δ)	of Com	pounds 2	-7 (in	CDCl ₃) ^a
----------	--------------------	-----------------	--------	----------	--------	----------------------------------

proton	2	3	4	4a	5	6	7
Η-1α	3.25 d	3.24 d	1.89 dd		3.75 dd	3.97 dd	1.99 dd
$H-1\beta$			2.60 dd				2.64 dd
$H-2\beta$	3.60 dd	3.60 dd	5.00 ddd	5.08 ddd	2.03 dd; 1.14 dd	2.01 dd; 1.16 dd	3.72 ddd
H-3a	3.15 d	3.14 d	3.20 d	4.74 d			3.02 d
$H-3\beta$					4.77 t	3.62 t	
H-6			5.20 dd	5.20 dd			
H-9	1.68 d	1.66 br d			1.78 d	1.77 d	1.72 br d
H-11 β	4.31 dd	4.36 dd	4.27 dd	5.45 dd	4.36 dd	4.35 dd	4.30 dd
H-12	5.27 d	5.21 d	5.17 d	5.17 d	5.25 d	5.24 d	5.17 d
CH-Me		0.86 d	0.85 d	0.88 d	0.88 d	0.84 d	0.84 d
		0.88 d	0.90 d	0.91 d	0.92 d	0.93 d	0.88 d
C-Me	0.83 s	0.70 s	0.80 s	0.79 s	0.80 s	0.80 s	0.79 s
	0.86 s	0.92 s	0.89 s	0.90 s	0.86 s	0.86 s	0.86 s
	0.88 s	1.03 s	1.04 s	0.91 s	0.91 s	0.95 s	0.92 s
	0.89 s	1.06 s	1.06 s	1.09 s	1.05 s	1.03 s	1.05 s
	0.98 s	1.15 s	1.15 s	1.19 s	1.10 s	1.08 s	1.06 s
	1.03 s	1.17 s	1.20 s	1.22 s	1.24 s	1.19 s	1.16 s
	1.10 s						
	1.21 s						
OAc			2.06 s	2.06 s; 1.93 s; 1.97 s	2.08 s		

^{*a*} J (Hz) = Compound **2**: 1 α ,2 β = 9; 2 β ,3 α = 10, 9,11 = 8.5; 11,12 = 3. Compound **3**: 1 α ,2 β = 9; 2 β ,3 α = 9.8, 9,11 = 9; 11,12 = 3; 19, 29 = 6.5; 20,30 = 6. Compound **4**: 1 α ,1 β = 13; 1 α ,2 β = 11; 1 β ,2 β = 5; 3 α ,2 β = 10; 6,7 α = 4; 6,7 β = 8; 9, 11 = 9; 11,12 = 3; 19, 29 = 6.5; 20,30 = 6. Compound **4a**: 1 α ,1 β = 13; 1 α ,2 β = 11; 1 β ,2 β = 5; 3 α ,2 β = 10; 6,7 α = 4; 6,7 β = 8; 9, 11 = 9; 11,12 = 3; 19, 29 = 6.5; 20,30 = 6. Compound **4a**: 1 α ,1 β = 13; 1 α ,2 β = 11; 1 β ,2 β = 5; 3 α ,2 β = 10; 6,7 α = 4; 6,7 β = 8; 9, 11 = 9; 11,12 = 3; 19, 29 = 6.5; 20,30 = 6. Compound **5**: 1 α ,2 β = 11; 1 α ,2 α = 5; 2 α ,3 β = 2 β ,3 β = 3;9,11 = 9; 11,12 = 3.5; 19, 29 = 6.5; 20,30 = 6. Compound **6**: 1 α ,2 β = 11.5; 1 α ,2 α = 5; 2 α ,3 β = 2 β ,3 β = 3;9,11 = 9; 11,12 = 3.5; 19, 29 = 6.5; 20,30 = 6. Compound **6**: 1 α ,2 β = 11; 1 β ,2 β = 4.5; 2 β ,3 α = 9.7.

NMR spectra. The HRMS of compound 2 indicated the molecular formula C₃₀H₅₀O₄ with six degrees of unsaturation, five of which were accounted for by a pentacyclic skeleton and one by a double bond. In the ¹H NMR spectrum, the presence of Δ , ¹²11-OH group, followed from signals at δ 5.27 (H-12) and 4.31 (H-11 $\hat{\beta}$). Similar compounds have been obtained from Salvia argentea.¹² The ¹³C NMR (APT) spectrum of 2 contained eight CH₃, seven CH₂, eight CH, and seven quaternary carbon signals (Table 2). Methyl signals at δ 0.83, 0.86, 0.88, 0.98, 1.04, 1.10, and 1.21 (each 3H, s) indicated an olean-type structure. Three successive secondary hydroxy groups followed from the signals at δ 3.25, 3.60 (H-2 β), and 3.15 (H-3 α). Spin decoupling experiments showed the C_1-C_3 and the $C_9 C_{12}$ sequences. Irradiation of H-2 (δ 3.60) collapsed the signals at δ 3.25 (H-1) and 3.15 (H-3) into singlets, while irradiation of H-11 (δ 4.31) collapsed the doublet signal of H-12 (δ 5.27) and a signal at δ 1.68 (J = 8.5 Hz) into singlets, the latter signal being assigned to H-9. Acetylation of 2 at room temperature yielded a known compound (11), first isolated from Salvia argentea,12 while drastic acetylation of **2** yielded the 2α , 3β , 11α -triacetyl derivative (**12**).¹² Thus, compound **2** was deduced to be 1β , 2α , 3β , 11α -tetrahydroxyolean-12-ene.

Compound **3** was the ursane analogue of compound **2**. The main difference in the ¹H NMR spectrum of **3** was observation of two methyl doublets at δ 0.86 and 0.88 assigned to an ursan skeleton and small chemical shift differences for H-11 and H-12. ¹H-¹H COSY experiments allowed assignment of ring A protons and ring C protons giving correlations among H-1 α , H-2 β , and H-3 α , as well as H-9 and H-11 and H-11 and H-12. Therefore, the structure of compound **3** was established as 1 β ,2 α ,3 β ,11 α -tetrahydroxy-urs-12-ene.

Compound 4, 2α -acetoxy-urs-5,12-diene- 3β ,11 α -diol, had the molecular formula $C_{32}H_{50}O_4$ (HRMS). A signal at δ 5.00 in the ¹H NMR spectrum of 4 indicated a hydrogen geminal to an acetyl group, with the acetyl signal at δ 2.06. This was also evident from ¹³C NMR signals at δ 167.8 and 21.2 (Table 2) and IR absorption bands at 1725 and 1270 cm⁻¹. The multiplicity (ddd) of the methine at δ 5.00 indicated

Table 2. ¹³C NMR Data (δ) of Compounds **2**–**7** (in CDCl₃)

С	2	3	4	5	6	7
1	76.5 d	76.8 d	44.2 t	76.1 d	75.8 d	38.6 t
2	75.1 d	75.0 d	73.2 d	27.2 t	28.3 t	69.2 d
3	79.7 d	79.9 d	71.3 d	70.4 d	72.1 d	82.4 d
4	38.2 s	38.2 s	40.4 s	38.2 s	38.2 s	38.1 s
5	54.6 d	54.6 d	144.9 s	55.1 d	55.2 d	55.2 d
6	18.1 t	18.0 t	121.7 d	18.0 t	18.1 t	18.3 t
7	31.3 t	31.3 t	37.3 t	32.6 t	32.7 t	33.1 t
8	41.4 s	41.3 s	40.6 s	40.2 s	40.2 s	40.4 s
9	55.0 d	54.6 d	54.3 d	54.3 d	54.2 d	54.5 d
10	43.6 s	43.8 s	38.6 s	38.2 s	38.3 s	37.8 s
11	69.5 d	69.2 d	70.1 d	70.2 d	70.3 d	70.1 d
12	123.5 d	123.4 d	124.6 d	124.5 d	124.6 d	124.2 d
13	145.1 s	144.2 s	145.3 s	145.2 s	145.3 s	144.9 s
14	44.6 s	44.5 s	44.7 s	44.6 s	44.5 s	43.8 s
15	27.7 t	27.8 t	27.6 t	27.5 t	27.6 t	28.0 t
16	27.8 t	27.9 t	27.5 t	27.4 t	27.5 t	27.2 t
17	32.2 s	32.8 s	32.6 s	32.7 s	32.7 s	33.0 s
18	40.3 d	60.1 d	59.8 d	59.9 d	60.0 d	59.7 d
19	46.2 t	41.8 d	41.6 d	40.3 d	40.2 d	39.8 d
20	31.1 s	41.8 d	41.6 d	41.6 d	41.5 d	40.9 d
21	33.7 t	30.9 t	31.0 t	31.0 t	31.1 t	30.8 t
22	32.4 t	41.5 t	41.4 t	41.6 t	41.7 t	41.5 t
Me's	28.2 q	28.1 q	28.3 q	28.2 q	28.2 q	28.1 q
	15.8 q	16.0 q	16.1 q	16.9 q	16.9 q	16.8 q
	15.7 q	16.1 q	20.7 q	15.9 q	16.0 q	16.1 q
	16.8 q	16.9 q	16.9 s	16.7 q	16.6 q	16.5 q
	27.4 q	23.3 q	24.4 q	23.5 q	23.6 q	23.6 q
	32.5 q	28.4 q	28.5 q	28.2 q	28.2 q	28.3 q
	24.6 q	17.5 q	17.8 q	17.1 q	17.2 q	17.3 q
~	21.4 q	21.4 q	21.5 q	21.2 q	21.5 q	21.6 q
OAc			21.2 q	21.0 q		
			167.8 s	170.7 s		

that it was between a methine and a methylene group. The signal at δ 3.20 and spin decoupling experiments showed the relationship between H-3 α and H-2 β . Irradiation of the signal at δ 3.20 (H-3) collapsed the signal at δ 5.00 (H-2) to a double doublet, while irradiation of H-2 collapsed H-3 to a singlet and collapsed H-1 β and H-1 α to doublets. Therefore, the acetyl group was clearly at C-2. Signals at δ 4.27 and 5.17 indicated an α -hydroxyl at C-11 and a double bond at C-12, as also observed in compounds **2** and **3**. A second double bond in the molecule was deduced from

the signal at δ 5.20. Considering the molecular formula and studying a Dreiding model, this double bond could be situated only between C-5 and C-6. Acetylation of 4 under reflux for 2 h yielded 4a; in the ¹H NMR spectrum, three acetyl groups were observed at δ 1.93, 1.97, and 2.06 (each 3H, s), the latter being originally present at C-2. The signal at δ 3.20 was shifted to 4.74 (H-3 α), the signal at δ 4.27 to 5.45 (H-11 β), and the signal at δ 5.00 was slightly shifted to 5.08 (H-2 β), although the olefinic proton signals at δ 5.17 (H-12) and 5.20 (H-6) were not affected. In 1D NOE experiments, irradiation of 5.08 (H-2 β) caused signal enhancements for C-10 Me and C-4 Me protons, indicating the stereochemistry of C-2 acetyl group to be α . HETCOR and the COLOC experiments were carried out with compound 4, and the methyl groups were assigned as follows: δ 1.20 (Me-24) and 0.89 (Me-25); other Me groups are at δ 0.80 (Me-26), 1.04 (Me-27), 1.06 (Me-28); 0.85 and 0.90 are assigned for Me-29 and Me-30. The HRMS and ¹³C NMR (APT) spectra of 4 (Table 2) were in agreement with the indicated structure.

Compound 5 (C32H52O4) had seven degrees of unsaturation, five of accounted for by a pentacyclic skeleton, a one by an acetyl carbonyl and one by a double bond. The ¹H NMR spectrum of 5 indicated an ursane-type structure (Table 1). COSY experiments clearly showed the relationships between nieghboring protons H-11 and H-12, as well as between H-11 and H-9. The signal at δ 3.75 (H-1 α) coupled with H-2 β (δ 2.03), the latter showing a cross-peak to the signal at δ 1.14 (H-2 α); H-3 β also showed cross-peaks to H-2 α and H-2 β . An acetyl methyl (δ 2.08) and the chemical shift of H-3 (δ 4.77) clearly indicated the presence of an acetyl group at C-3; its α position was deduced from its splitting pattern and comparison with similar compounds having a β -hydroxy or -acetoxy group at C-3. 1β , 3β ,-11α-Trihydroxy-olean-12-ene (10) was isolated from Maytenus horrida,13 and its 3-acetoxy derivative has been prepared (13), the differences between compounds 5 and 13 were in the chemical shift and splitting pattern of H-3, as well as in the presence of two methyl doublets (Me-29 and Me-30). The ¹³C NMR (APT) spectrum of 5 revealed nine CH_3 , seven CH_2 , nine CH, and seven guaternary signals for 32 carbon atoms of the molecule. Thus, the structure of 5 was established as 3α-acetoxy-urs-12-ene- 1β , 11α -diol.

The ¹H NMR spectrum of 6 indicated an ursane triterpene. A triplet signal at δ 3.62 (J = 3.5 Hz) was very characteristic for a C-3 β proton, indicating that it bears an α -hydroxy group. In fact, ¹H NMR signals were very similar to those of compound 5, the only meaningful difference was observed for H-3 β (δ 3.62). The chemical shift difference in H-3 α compared to that of 5 was attributed to the presence of a hydroxy group at C-3 instead of an acetyl group in compound 6. This was proven by the lack of an acetyl methyl signal. The HRMS spectrum showed a molecular ion peak at m/z 458.3660, in the EIMS; the loss of the first hydroxy group at m/z 440 and the second at m/z 422 was clearly observed. Deacetylation of 5 yielded compound 6, verifying its structure. Thus, the structure of compound **6** was established as 1β , 3α , 11α trihydroxy-urs-12-ene.

The ¹H NMR spectrum of **7** also indicated an ursane skeleton. Two methine proton signals were observed at δ 3.02 and 3.72, assigned to H-3 α and H-2 β , respectively. Based on spin-decoupling experiments H-1 β and H-1 α signals were assigned to signals δ 2.64 and 1.99, respectively. The Δ^{12} ,11-OH moiety was observed in δ 4.30 (dd, J = 3.2, 10.0 Hz, H-11 β) and 5.17 (d, J = 3.2 Hz, H-12) as

followed in all the above triterpenoids. A similar compound, 2α , 3β -dihydroxy-urs-12-ene¹⁴ without the C-11 hydroxy group, had a similar spectrum. The HRMS of **7** gave a molecular ion peak at m/z 458.3554, and the ¹³C NMR spectrum of **7** was consistent with the assigned structure. Thus, **7** was established as 2α , 3β , 11α -trihydroxy-urs-12-ene.

Experimental Section

General Experimental Procedures. EIMS and HRMS were recorded on a ZabSpec mass (micromass) spectrometer. ¹H and ¹³C NMR were run on a Bruker Ac 200 L NMR spectrometer at 200 and 50.32 MHz, respectively, in CDCl₃. UV spectra were taken on a Varian Techtrone 535, and IR, on a Perkin–Elmer 983 instrument. Si gel 60 (70–230 mesh, E. Merck) was used for column chromatography, preparative TLC was performed on Si gel (UV₂₅₄ precoated) plates.

Plant Material. The roots of *Salvia kronenburgii* were collected from eastern Turkey (Van) in June 1997, at 2250 m altitude and identified by Prof. Dr. M. Koyuncu and Dr. Nasip Demirkus. A voucher specimen is deposited in the Herbarium of 100. Yil University, Van (Demirkuş 5515 VANF).

Extraction and Isolation. The powdered plant material (2.4 kg) was macerated with Me₂CO at room temperature, yielding 30 g of a residue. The residue was roughly separated on a Si gel column (5 \times 110 cm) eluting with hexane and gradients of CHCl₃ and MeOH. The similar fractions were combined to give five main fractions (A-E). Fraction A (1-20), having only the fatty materials, was discarded. Fraction B (21–29) (220 mg) was subjected to a smaller Si gel column $(2 \times 70 \text{ cm})$ eluting with CHCl₃ to give known diterpenoids taxodione (95 mg), ferruginol (55 mg), salvipisone (22 mg), and 12-hydroxysapriparaquinone (15 mg) and the new diterpenoid kronenquinone (1). From fraction C (30-48) (800 mg) on a Si gel column (4 \times 100 cm), eluting with CHCl₃-Me₂CO (95:5), acetoxyroyleanone (45 mg), royleanone (25 mg), 1-oxosalvibretol (17 mg), and Δ^7 -manool (14 mg) were obtained. Fraction D (49–74) (440 mg) was also subjected to a Si gel column (3 \times 80 cm) and eluted by $CHCl_3$ and increasing amounts of MeOH to afford 6-hydroxysalvinolone (13 mg) and the new triterpenoids 2 (15 mg), 3 (13 mg), 4 (15 mg), 5 (18 mg), 6 (20 mg), and 7(14 mg). The known compounds were identified by comparison with literature data.

Kronenquinone (1): UV (CHCl₃) λ_{max} (log ϵ) 430 (3.0), 395 (3.2), 282 (4.1) nm; IR (CHCl₃) ν_{max} 3480, 3050, 2960, 2870, 1675, 1640, 1620, 1595, 1505, 1460, 1380, 950, 880 cm⁻¹; ¹H NMR (CDCl₃) δ 7.64 (1H, d, J = 9 Hz, H-7), 7.18 (1H, d, J =9 Hz, H-6), 3.38 (1H, sept, J = 7 Hz, H-15), 2.65 (3H, s, Me-18), 2.54 (3H, s, Me-19), 1.33 (3H, d, *J* = 7 Hz) and 1.32 (3H, d, J = 7 Hz) (Me-16 and Me-17); ¹³C NMR (CDCl₃) δ C-1 153.2 s, C-2 135.7 s, C-3 141.7 s, C-4 133.4 s, C-5 127.6 s, C-6 122.8 d, C-7 133.5 d, C-8 126.0 s, C-9 127.3 s, C-10 150.1 s, C-11 182.8 s, C-12 150.5 s, C-13 134.9 s, C-14 184.2 s, C-15 26.2 d, C-16 22.8 q, C-17 22.7 q, C-18 21.1 q, C-19 17.7 q; EIMS m/z(rel int) 326 [M]⁺ (7), 312 (100), 300 (12), 281 (32), 256 (60), 243 (88), 227 (77), 219 (20), 213 (21), 211 (21), 203 (11), 197 (13), 185 (19), 165 (17), 155 (12), 149 (20), 141 (16), 128 (17), 123 (11), 109 (12), 95 (11), 83 (20), 71 (62), 57 (21); HREIMS m/z 326.1136 (calcd for C₁₉H₁₈O₅, 326.1144).

1 β ,**2** α ,**3** β ,**11** α -**Tetrahydroxy-olean-12-ene** (**2**): $[\alpha]^{25}_{D} 0.0^{\circ}$ (*c* 0.1, CHCl₃); IR (CHCl₃) ν_{max} 3450, 3380, 2960, 2850, 1645, 1380, 1260, 1100, 1080, 1050, 980 cm⁻¹; ¹H and ¹³C NMR, Tables 1 and 2; EIMS *m*/*z* (rel int) 474 [M]⁺ (15), 456 [M - H₂O]⁺ (100), 440 [M - 2×H₂O]⁺ (17), 423 [M - 2×H₂O - OH]⁺ (14), 405 [M - 2×H₂O - 2×OH]⁺ (16), 324 (5), 303 (16), 285 (7), 267 (5), 255 (23), 189 (6), 171 (8), 119 (7), 107 (6), 95 (5); HREIMS *m*/*z* 474.3698 (calcd for C₃₀H₅₀O₄, 474.3710).

1 β ,**2** α ,**3** β ,**11** α -**Tetrahydroxy-urs- 12-ene (3)**: IR(CHCl₃) ν_{max} 3455, 3380, 2960, 2850, 1645, 1380, 1255, 1100, 1080, 1050, 975 cm⁻¹; ¹H and ¹³C NMR, Tables 1 and 2; EIMS *m*/*z* (rel int) 474 [M]⁺ (12), 456 [M - H₂O]⁺ (100), 440 [M - $2 \times H_2O$]⁺ (15), 423 [M - $2 \times H_2O$ - OH]⁺ (10), 405 [M - $2 \times H_2O$ - $2 \times OH$]⁺ (15), 324 (5), 302 (18), 284 (6), 266 (4), 255 (20), 189 (8), 171 (9), 107 (9), 95 (7); HREIMS m/z 474.3696 (calcd for C₃₀H₅₀O₄, 474.3710).

2α-**Acetoxy-urs-5,12-diene-3**β,11α-**diol (4)**: $[\alpha]^{25}{}_{\rm D} - 50.0^{\circ}$ (*c* 0.1, CHCl₃); IR (CHCl₃) $\nu_{\rm max}$ 3460, 3050, 2980, 2862, 1725, 1640, 1605, 1360, 1270, 880 cm⁻¹; ¹H and ¹³C NMR, Tables 1 and 2; EIMS *m*/*z* (rel int) 498 [M]⁺ (27), 482 [M - 16]⁺ (100), 467 [M - 16 - CH₃]⁺ (18), 441.2 [M - OCOCH₃]⁺ (72), 423 (20), 407 (19), 389 (17), 329 (20), 269 (21), 255 (45), 234 (34), 218 (15), 204 (20), 191 (22), 171 (14), 145 (18), 133 (25), 123 (25), 109 (28), 95 (35), 69 (32), 57 (22), 43 (24); HREIMS *m*/*z* 498.3664 (calcd for C₃₂H₅₀O₄, 498.3710).

Acetylation of 2, 4, and 10. Of each compound (2, 4, and 10) 5 mg was dissolved in 1 mL of pyridine, and 1 mL Ac₂O was added. The reaction mixtures were left at room temperature for 36 h, evaporated to dryness, and the residues subjected to preparative TLC using $CHCl_2$ to give 11, 4a, and 13.

Drastic Acetylation of Compound 2. Of compound **2** 5 mg dissolved in 1 mL of pyridine, and 1 mL Ac_2O was added. The mixture was refluxed in a water bath for 2 h, left at room temperature for 24 h, and evaporated to dryness to afford compound **12**.

Urs-5,12-diene-2α,**3**β,**11**α-**triacetate (4a):** IR (CHCl₃) ν_{max} 2862, 1730, 1722, 1640, 1600, 1360, 1280, 880 cm⁻¹; ¹H NMR, Table 1; HREIMS *m*/*z* 582.3930 (calcd for C₃₆H₅₄O₆, 582.3920).

3α-**Acetoxy-urs-12-ene-1**β,**2**α-**diol** (5): IR (CHCl₃) ν_{max} 3440, 3055, 2960, 1727, 1640, 1380, 1265, 890 cm⁻¹; ¹H and ¹³C NMR, Tables 1 and 2; EIMS (70 eV) *m/z* (rel int) 500 [M] + (71), 482 [M - H₂O]⁺ (100), 464 [M - 2×H₂O]⁺ (8), 455 (10), 422 [482 - AcOH]⁺ (65), 406 (35), 389 (33), 329 (11), 288 (12), 273 (14), 255 (30), 217 (82), 189 (72), 175 (62), 135 (45), 109 (36), 95 (51), 81 (20), 69 (26); HREIMS *m/z* 500.3800 (calcd for C₃₂H₅₂O₄, 500.3865).

1 β ,**3** α ,**11** α -**Trihydroxy-urs-12-ene (6):** [α]²⁵_D 0.0° (*c* 0.1, CHCl₃); IR (CHCl₃) ν_{max} 3450, 3380, 2960, 2850, 1645, 1380, 1255, 1100, 1080, 1050, 975 cm⁻¹; ¹H and ¹³C NMR, Tables 1 and 2; EIMS (70 eV) *m*/*z* (rel int) 458.3 [M]⁺ (70), 440.3 [M - H₂O]⁺ (90), 425.3 [M - H₂O - CH₃]⁺ (21), 422.3 [M - 2×H₂O]⁺ (21) 407.2 [M - 2×H₂O - CH₃]⁺ (22), 389.3 (23), 355.2 (7),

288.2 (12), 273.2 (17), 255.2 (40), 247.2 (63), 235.1 (33), 217.1 (45), 203.1 (38), 193.1 (100), 189.1 (72), 175.1 (64), 135.1 (47), 121.1 (56), 108.9 (45), 95.1 (53), 81.0 (23), 69.0 (19); HREIMS m/z 458.3660 (calcd for $C_{30}H_{50}O_3$, 458.3759).

2 α ,3 β ,11 α -**Trihydroxy-urs-12-ene (7):** IR (CHCl₃) ν_{max} 3460, 3380, 2960, 2855, 1640, 1380, 1260, 1100, 1075, 1050, 975 cm⁻¹; ¹H and ¹³C NMR, Tables 1 and 2; EIMS (70 eV) *m/z* (rel int) 458 [M]⁺ (15), 442 [M - 16]⁺ (100), 424 [442 - H₂O]⁺ (83), 409 [424 - Me]⁺ (17), 355 (18), 322 (10), 271 (22), 255 (25), 203 (17), 135 (15), 108 (20), 95 (23), 78 (62), 69 (45); HREIMS *m/z* 458.3554 (calcd for C₃₀H₅₀O₃, 458.3759).

Acknowledgment. This study was supported by the Research Fund of the University of Istanbul, project no. 1341/2800799, given to one of us (G. Topçu).

References and Notes

- Ulubelen, A.; Topçu, G. In *Studies in Natural Products Chemistry*; Atta-ur-Rahman, Ed.; Elsevier: New York, 1998; Vol. 20, pp 659– 718.
- (2) Ulubelen, A.; Topçu, G.; Bozok-Johansson, C. J. Nat. Prod. 1997, 73, 1275–1280.
- (3) Topçu, G.; Kartal, M.; Ulubelen, A. Phytochemistry 1997, 44, 1393– 1395.
- (4) Topçu, G.; Ulubelen, A. J. Nat. Prod. 1996, 59, 734-737.
- (5) Topçu, G.; Eri, C.; Ulubelen, A. *Phytochemisty* **1996**, *41*, 1143–1147.
 (6) Kupchan, S. M.; Karim A.; Marcks, C. J. Am. Chem. Soc. **1968**, *90*, 5923–5924.
- (7) Cambie, R. C.; Madden, R. J.; Parnelly, J. C. Aust. J. Chem. 1971, 24, 217–221.
- (8) Michavila, A.; de la Torre, M. C.; Rodriguez, B. *Phytochemistry* 1986, 25, 1935–1937.
- (9) Rüedi, P. Helv. Chim. Acta 1984, 67, 1116-1120.
- (10) Topçu, G.; Ulubelen, A. J. Nat. Prod. **1996**, *59*, 734–737.
- (11) Bohlman, F.; Zdero, C.; Hoffmann, E.; Mahanta, P. K.; Dorner, W. *Phytochemistry* **1987**, *17*, 1917–1922.
 (12) Gonzalez, A. G.; Ferro, E. A.; Ravela, A. G. *Phytochemistry* **1987**, *26*,
- (12) Contaction, A. G., Phys. Rev. A, Rev. C. Phytochemistry 1001, 20, 2785–2788.
 (13) Bruno, M.; Savona, G.; Hueso-Rodriguez, J. A.; Pascual, C.; Rodriguez,
- B. *Phytochemistry* **1987**, *26*, 497–501. (14) Eggert, H.; Van Antwerp, C. L.; Bhacca, N. S.; Djerassi, C. J. Org.
- (14) Eggert, H.; Van Antwerp, C. L.; Bhacca, N. S.; Djerassi, C. J. Org. Chem. **1976**, 41, 71–78.

NP990165P